

Syntheses of 12-Oxygenated Forskolins

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The preparation of selectively oxygenated forskolin derivatives has been achieved with reagent, solvent, and steric environment as key factors in selectivity of the oxidation.

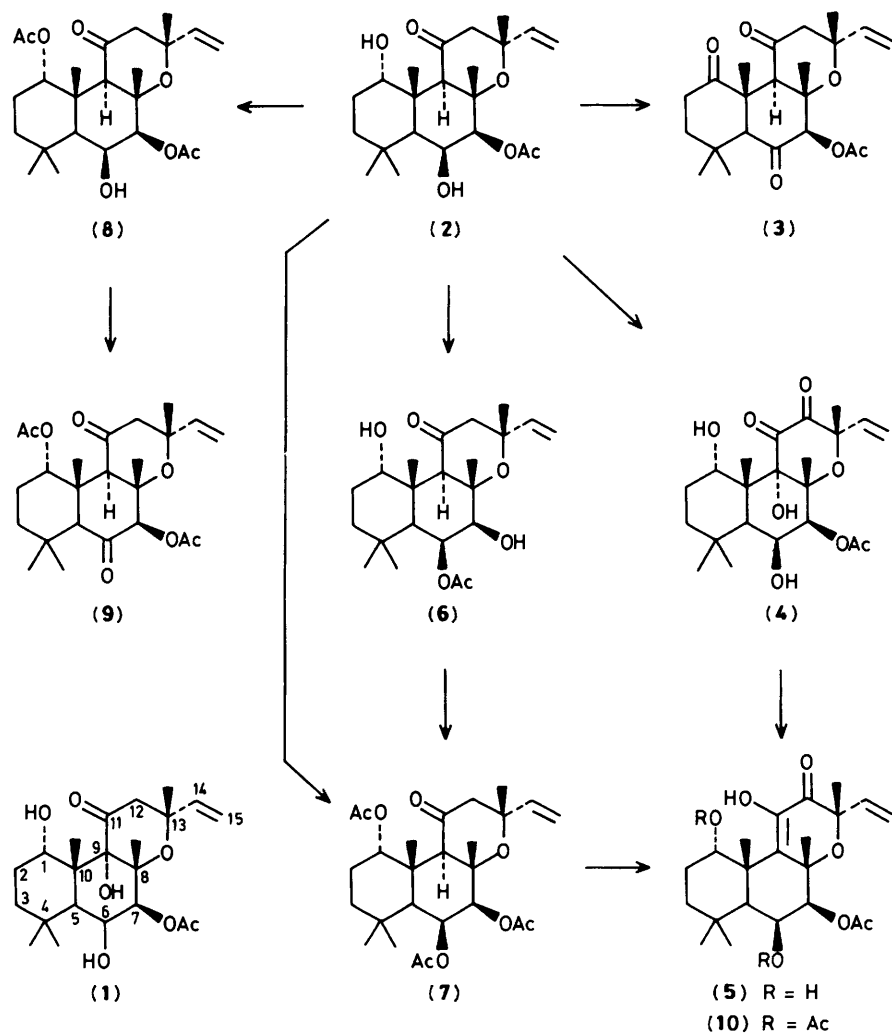
Forskolin (**1**), a biologically active diterpene isolated from *Coleus forskohlii*, has generated considerable interest as a target for total or partial synthesis of the natural product itself and of potentially active analogues as well.^{1,2}

We described in a previous paper the successful conversion of 9-deoxyforskolin (**2**) into forskolin *via* a stereo- and regio-selective hydroxylation sequence.³ We were also interested in developing procedures which would allow direct hydroxylation at C-9 with or without further functionalization of the forskolin framework. This paper describes some of the results of these studies, including the preparation of the previously unknown 12-oxygenated derivatives of both forskolin and 9-deoxyforskolin.

The carbon framework of 9-deoxyforskolin proved surprisingly resistant to a number of oxidizing agents such as lead tetra-acetate,⁴ molecular oxygen with various catalysts,⁵ or halogenating agents.⁶ Benzeneseleninic anhydride has been reported to effect α -hydroxylation of ketones in the presence of strong base.⁷ However, under these conditions [(PhSeO)₂O, NaH, toluene, reflux] the only product we could

obtain from 9-deoxyforskolin was the triketone (**3**) (40%).[†] The utilization of selenium dioxide⁸ as an oxidizing agent

[†] Selected spectral data: (**3**) i.r. (CHCl₃): ν_{\max} 1735s and 1715s cm⁻¹; ¹H n.m.r. (Varian XL-200; CDCl₃): δ 6.08 (1H, dd, *J* 12 and 18 Hz, 14-H), 5.30 (3H, m, 2 \times 15-H, 7-H), 3.92 (1H, s, 5-H), 3.15 (1H, dt, *J* 4 and 10 Hz), 2.95 (1H, *J*_{AB} 18 Hz, 12-H), 2.58 (1H, *J*_{AB} 18 Hz, 12-H), 2.25 (3H, s, Ac), 2.20 (1H, m), 1.50 (3H, s, Me), 1.45 (3H, s, Me), 1.35 (3H, s, Me), 1.24 (3H, s, Me), 1.00 (3H, s, Me), and 1.84–1.2 (2H, m); *m/z* 390 (*M*⁺, EI, 17 eV); m.p. 127–129°C; (**4**) i.r. (CHCl₃): ν_{\max} 1740s, and 1730s cm⁻¹; ¹H n.m.r. (CDCl₃) δ 7.40 (1H, s, 1-OH), 6.00 (1H, dd, *J* 12 and 18 Hz, 14-H), 5.55 (2H, m, 15-, 7-H), 5.18 (1H, dd, *J* 2 and 12 Hz, 15-H), 4.68 (1H, m, 1-H), 4.50 (1H, m, 6-H), 2.20 (3H, s, Ac), 1.74 (3H, s, Me), 1.54 (3H, s, Me), 1.28 (3H, s, Me), 1.07 (3H, s, Me), and 2.4–1.0 (7H, m); *m/z* 406 (*M*⁺ – 18, 17 eV); m.p. 185–187°C (decomp.); (**5**) i.r. (CHCl₃): ν_{\max} 1742s and 1665m cm⁻¹; ¹H n.m.r. (CDCl₃): δ 5.90 (1H, dd, *J* 9 and 18 Hz, 14-H), 5.30 (3H, m, 2 \times 15-H, 7-H), 4.70 (1H, m, 1-H), 4.58 (1H, m, 6-H), 2.22 (3H, s, Ac), 1.91 (3H, s, Me), 1.6 (3H, s, Me), 1.57 (3H, s, Me), 1.28 (3H, s, Me), 1.06 (3H, s, Me), and 2.30–1.0 (m, 8H); *m/z* 408 (*M*⁺, 17 eV); m.p. 89–91°C.



provided us with the first clear evidence of functionalization at the saturated C-9 and C-12 positions. Thus, treatment of 9-deoxyforskolin with this reagent (SeO_2 , pyridine, toluene, reflux) proceeded cleanly, affording as the only product 12-oxoforskolin (4) (91%). The choice of solvent was critical in this case; no reaction occurred utilizing toluene, tetrahydrofuran, or dioxane alone.

In addition to standard ^1H n.m.r., i.r., mass spectral, and elemental analyses, \dagger 12-oxoforskolin (4) was characterized by its ^{13}C n.m.r. spectrum which showed two ketone carbonyl carbon signals (δ 199.2 and 186.3) as well as a signal attributed to an ester carbonyl (δ 169.6).

Interestingly, forskolin itself was unstable under these conditions, suggesting that C-12 might be the site of initial oxidation. This hypothesis is supported by the observation that reduction of (4) (Zn , HOAc , room temp.) provided as sole product 12-oxo-9-deoxyforskolin (5) \dagger in 94% yield. The ^{13}C n.m.r. spectrum of this compound revealed that the 11-ketone was completely enolized (one ketone carbonyl carbon signal at δ 193.6, two new sp^2 carbon signals at δ 141.2 and 137.3). These two compounds comprise the first examples of 12-oxygenated forskolin derivatives.

In the course of these studies we also prepared a series of selectively protected acetyl derivatives of 9-deoxyforskolin, which were also subjected to oxidative conditions. Since the

6-acetyl-7-deacetyl derivative (6) is not accessible *via* direct acylation, treatment of (2) with base [$\text{LiN}(\text{SiMe}_3)_2$, tetrahydrofuran, 0°C] 3 resulted in a clean 7 \rightarrow 6 acyl migration to provide (6) (80%). 9 Exposure of this compound to acetic anhydride (Ac_2O , pyridine, reflux) produced the fully acetylated derivative (7) (75%), while under identical conditions 9-deoxyforskolin gave only the 1,7-diacetyl derivative (8) (87%). Alternatively, (7) could be prepared directly from (2) under acidic conditions (Ac_2O , cat. HClO_4 , 85%).

While exposure of the 1,7-diacetyl derivative (8) to benzeneseleninic anhydride [$(\text{PhSeO})_2\text{O}$, NaH , toluene, reflux] provided the diketone (9) (47%), under these conditions the fully protected derivative (7) gave a complex mixture of products. However, treatment of (7) with selenium dioxide (SeO_2 , pyridine, reflux) provided the enolic ketone (10) (84%) as sole product, the result of oxidation at the 12-position only. Apparently, the presence of an acetate moiety in the 1-position is sufficient to render the already hindered 9-position inaccessible to further oxidative attack.

In conclusion, we have developed methodology to prepare the hitherto unknown 12-oxygenated derivatives of forskolin with or without concomitant functionalization at the 9-position. The biological activity of these new compounds is currently under evaluation.

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